Using ISSR-Markers for Genetic Diversity of some Representatives of Orchidaceae

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The present study is directed to determination the genetic diversity of representatives of *Orchidaceae* family using six inter simple sequence repeat (ISSR) primers. The possibility of using them to assess the level of genetic polymorphism is shown. Monomorphic and polymorphic bands indicating the species and genus determination have been allocated which enables to use them for making genetic formulas and certification of populations. Cluster analyses on the base of UPGMA (unweighted pair group method with arithmetic mean) made it possible to estimate the degree of similarity of the tested species.

Key words: Genetic diversity, Orchidaceae, ISSR markers.

The *Orchidaceae* is one of the largest angiosperm families^{1,2,3,4}. It lists about 750 genera and 20,000-25,000 species⁵. Most orchid species are found in the tropical regions¹. Bryansk region is a unique floristic region, located in an intersection of important natural, botanical and geographical boundaries. This is an important factor of the high level of biodiversity in the region.

The Red Book of the Bryansk region was written in 2004. An electronic, constantly updated database "Rare species of Sudost and Desna interfluve", containing more than 2,000 entries about location of rare species of the Bryansk region and their phytocenotic confinement was created which became the basis for monitoring the status of their cenopopulations. There are 18 genera and

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35 species of orchids in Bryansk region at present⁶.

Various methods are available for assessing the condition of populations of rare and endangered plant species. One of the main methods is determination of the level of genetic polymorphism. Inter simple sequence repeats (ISSR) were reported to be a reliable method to determine genetic polymorphism, due to the presence of a large number of microsatellite repeats in the genome of plants. This method of molecular analysis was suggested by Zietkiewicz et al. in 1994 and is based on variations between microsatellites found in the regions^{7.8}.

We have chosen the ISSR method to conduct the molecular genetic analysis⁹.

MATERIALSAND METHODS

We used DNA of rare and endangered species, collected in the six regions of the Russian Federation: Bryansk, Kaluga, Kursk, Orel, Smolensk and Tula (*Epipactis helleborine* (L.) Crantz., *Epipactis atrorubens* (Hoffm.) Besser, *Goodyera repens* (L.) R. Br., *Platanthera bifolia* L., *Ñypripedium calceolus* L., *Neottia nidus-avis* (L.) Rich., *Dactylorhiza incarnata* (L.) Soo., *Dactylorhiza maculate* (L.) Soo., *Digitalis grandiflora* Mill., *Lílium martagon* L., *Iris aphylla* L.). DNA of the three tropical species of Bedugul's Botanical Gardens collection (Indonesia) (*Eria multiflora* (Blume) Lindl., *Dendrobium malbrownii* Dockrill and *Pholidota gibbosa* (Blume) Lindl. ex de Vriese) was also extracted.

DNA was extracted from young leaves and dry leaf tissues at different time using the Cetyl trimethyl ammonium bromide (CTAB) method.

Six primers were selected for each species tested: IS1((GÀ)7 G(Y)G), IS2((AC)8G), IS3 ((GA)8C), IS4 ((CA)8A), IS5 ((CA)7(R)C) è IS6 ((AG)7(Y)T).

Each 20 μ l of the reaction contained 6 μ l of genomic DNA, 1 unit *Taq* DNA polymerase, 2,5 μ l dNTP, 90 pmol of each primer, 2 μ l PCR reaction buffer and water. The initial denaturation step at 94 °C for 5 min was followed by 40 cycles of amplification: 94 °C for 45 s, annealing for 45 s, and extension at 72 °C for 1,3 min. The final cycle at 72 °C for 7 min. The temperature of fusion was chosen experimentally.

Extraction of DNA and PCR was performed in 1-3 repeats, due to the rarity of the studied plants and the lack of ability to get enough DNA.

The PCR products were electrophoresed in a 2% agarose gels and stained with ethidium bromide. ISSR amplification products were scanned using GelDocXR system (BioRad, USA) and program for processing electrophoregrams Quantity One. To determine the size of the DNA fragments M27 marker (Sibenzime, Moscow) was used.

Data analysis

Molecular genetic certification was carried out on the basis of the approach proposed by Bronnikova for natural populations of representatives of the family *Campanulaceae* Juss.: *Adenophora lilifolia* (L.) A.DC.¹⁰.

The study included collecting herbarium material, choosing ISSR markers, PCR analysis, processing electrophoregrams and preparation of certification.

RESULTS AND DISCUSSION

Analyzed electrophoregrams revealed monomorphic bands which allows to attribute samples to genus or species and polymorphic bands specific to individual populations (Fig. 1).

Based on this data a type of formulas indicating fragments common to all samples of the studying population and the type of the fragment (g (genus), s (species) and p (polymorphic)) was composed (Table 1). At the beginning of the formula there goes an abbreviation indicating genus in capital letters and small letter indicates species, subscript specifies the type of the fragment, the length of the fragment and the subscript marker used. So the molecular genetic formula for Dactylorhiza maculate with primer IS4, can be written as DTm_s535₁₅₄. In order to determine whether these fragments belong to the species or taxa of higher rank, the results of molecular genetic analysis of the species belonging to different families were compared

Further, these formulas can be used to compile genetic passports and barcode of populations.

In species of genus *Platanthera* a total of 2901 DNA fragments were obtained with 6 primers. The highest number of bands (28) was obtained with IS1 primer. Monomorphic bands were not detected. However, with the IS1 primer, 1176 bp band was identified, it is found only in the population of the Bryansk region, Diatkovo area.

Despite the absence of monomorphic bands, there can be identified bands found in most samples. So with primer IS1 a fragment of 200 bp was found in 85%, in 82.5% - of 220 bp, 80% - 280 bp, 57.5% - 330 and 305 bp., with primer IS2 in 55% - 230 bp, with primer IS3 in 87.5% - 200 bp, with primer IS5 in 51.2% - 370 bp and primer IS6 in 92,1% - 160 and 190 bp to 68.4% - 220 bp.

We suppose that the absence of monomorphic bands in studied representatives of the genus *Platanthera* is connected with high levels of genetic diversity. To obtain more perfect data, it is necessary to collect more plant material from much bigger population, which is difficult due to using herbarium material, rarity of the samples and necessity to save natural populations.

Population	Number of samples	Symbol of population	Type of fragment	Molecular genetic formula
Cypripedium	8	СР	genus (g)	$CP_{g}493_{ISS}; CP_{g}460_{ISS}; CP_{g}418_{ISS}; CP_{g}361_{ISS}; CP_{g}300_{ISS}$
			species (s)	$CPm_s 401_{1S5}$
			polymorphic (p)	-
Dactylorhiza	12	DT	genus (g)	$DT_{g}190_{IS1}; DT_{g}290_{IS3}; DT_{g}180_{IS4};$
				$DT_{g}^{3}37_{IS6}^{}; DT_{g}^{2}266_{IS6}^{}; DT_{g}^{2}245_{IS6}^{};$
				$DT_{g}^{2}218_{IS6}; DT_{g}^{2}125_{IS6}$
			species (s)	$DTm_{s}535_{1S4}; DTm_{s}340_{1S4}; DTm_{s}84_{1S6}$
			polymorphic (p)	-
Epipactis	12	EP	genus (g)	$EP_{g}60_{IS1}$; $EP_{g}208_{IS5}$; $EP_{g}100_{IS6}$
			species (s)	-
			polymorphic (p)	$EPh_{p}532_{IS5}; EPh_{p}440_{IS5};$
Platanthera	41	PL	genus (g)	-
			species (s)	-
			polymorphic (p)	$PLb_{p}1176_{IS1}$
Tropicalorchids	3	ERm	genus (g)	-
		DNm	species (s)	-
		PHg	polymorphic (p)	$64_{154}; 300_{156}$
Neottia	12	NT	genus (g)	-
			species (s)	-
			polymorphic (p)	-

 Table 1. Molecular genetic certification of populations

Further, these formulas can be used to compile genetic passports and barcode of populations.

When the study samples of genus *Neottia* monomorphic bands also weren't detected. In 54.5% of cases with primer IS1 297 bp and 172 bp bands were identified, in 66.7% - 272 bp and 230 bp, in 58.3% - 320 bp, with primer IS2 in 58,3% - 355

bp., 50% - 500 bp and with primer IS3 in 58,3% - 233 bp.

In order to select extragenic bands and bands of a higher rank DNA from the samples of the families *Liliaceae* (*Lilium martagon*),

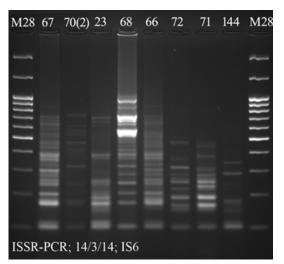


Fig. 1. ISSR patterns of genus *Epipactis* amplified with primers IS6

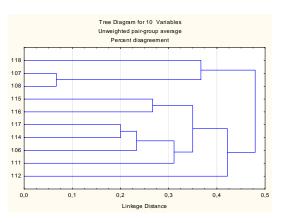


Fig. 2. UPGMA dendrogram of genetic distances among populations of genus *Dactylorhiza* with IS4 primer

Plantaginaceae (Digitalis grandiflora), Orchidaceae (Goodyera repens) and tropical orchids (Eria multiflora, Dendrobium malbrownii and Pholidota gibbosa) was extracted

In the course of the experiment, we could not find monomorphic bands common to members of all families. However, a fragment 64 bp with IS4 primer and a fragment 300 bp with primer IS6, were found in all samples of tropical orchids. Due to the small number of samples, we cannot assume the relationship with the habitat or location.

Similarities were calculated with Statistica 6.0 computer program and the cluster analysis was performed to develop a dendrogram by unweighted pair group method with arithmetic mean (UPGMA).

The closest accordance data of genetic relationship with location of populations obtained with primers IS4 and IS5 (Fig. 2).

According to this dendrogram samples of *Dactylorhiza incarnata*: 115 and 116, 114 and 117 are combined in the common clusters that match to their populations of Bryansk region, Sevskii district. Samples *Dactylorhiza maculate* 107 and 108 are also combined into a common cluster and belong to the population of Kaluga region. On the dendrogram these samples are at the greatest genetic distance from others, and in fact the population is most distant from the others, and most close to it is a sample of *Dactylorhiza incarnata* 118 (Bryansk region, Diatkovo district), which borders with Kaluga region, which is also shows in the dendrogram.

CONCLUSIONS

The results of molecular genetic analysis of representatives of *Orchidaceae* reveal the possibility of using ISSR-primers for analysis of intra- and interspecies differences. The results suggested that the genus *Platanthera* and *Neottia* are characterized by the highest level of genetic polymorphism.

Combinations of polymorphic DNA fragments that are specific for the studied genera, species or populations, can be recommend for their genetic certification.

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