# **Phage Display Technique and Hepatitis Viruses Studies**

# Majid Asadi Ghalehni<sup>1\*</sup> and S.M. Alavian<sup>1,2</sup>

<sup>1</sup>Gastroenterology and Liver Disease Research Center, Baqiyatallah University of Medical Sciences, Tehran, Iran. <sup>2</sup>Middle East Liver Diseases (MELD) Center, Tehran, Iran.

http://dx.doi.org/10.13005/bbra/2315

(Received: 14 May 2016; accepted: 15 July 2016)

Phage display is an in vitro selection method in which a peptide or protein is expressed as a fusion with a coat protein of a bacteriophage. This versatile technique has a lot of application in life science. In this literature review the applications of phage display in Hepatitis studies evaluated. This is the first review that considers this issue.

Keywords: Phage display, Hepatitis viruses.

Hepatitis is a medical condition defined by the inflammation of the liver and characterized by the presence of inflammatory cells in the tissue of the organ. Hepatitis may occur with limited or no symptoms, but often leads to jaundice (a yellow discoloration of the skin, mucous membrane, and conjunctiva), poor appetite, and malaise. Hepatitis is acute when it lasts less than six months and chronic when it persists longer. Acute hepatitis can be self-limiting (healing on its own), can progress to chronic hepatitis, or, rarely, can cause acute liver failure<sup>1</sup>.

Viral hepatitis is a major public health problem worldwide. The infection is caused by five taxonomically unrelated human viruses, namely, hepatitis A virus (HAV), hepatitis B virus (HBV), hepatitis C virus (HCV), hepatitis D virus (HDV) and hepatitis E virus (HEV).

HAV or hepatitis A virus cause an acute and commonly self-limiting infection followed by long immune protection against the virus<sup>2</sup>.

HBV, responsible for 600000 deaths each year, caused a life threatening liver disease. The

infectious virus particle, also called as Dane particle, is responsible for causing infection in 5 percent of world's population with 2 billion people infected with the virus and 350 million as carrier of chronic infection<sup>3-5</sup>.

Hepatitis C virus (HCV) is the major causative agent of non-A, non-B hepatitis worldwide. HCV is blood borne pathogen which causes severe liver disorders, including hepatocellular carcinoma, hepatic steatosis, liver cirrhosis, end stage liver disease and various metabolic disorders. HCV was identified by Choo et al. as a positive stranded RNA molecule related to Togaviridae or Flaviviridae<sup>6-7</sup>.

HDV is a unique agent characterized by a single-stranded RNA genome encapsidated by the hepatitis B surface antigen (HBsAg) and a peculiar strategy of infection of the target organ<sup>8</sup>. In fact, HDV requires the helper functions provided by hepatitis B virus (HBV) in order to propagate to hepatocytes, it can only infect subjects with co-existing HBV infection due either to the simultaneous transmission of the two viruses or super infection in an established HBV carrier<sup>9-10</sup>.

HEV is the sole member of the genus Hepevirus in the family of Hepeviridae, is the major cause of waterborne hepatitis in tropical and subtropical countries and of sporadic cases of viral

<sup>\*</sup> To whom all correspondence should be addressed. Tel.: +989128382915; Fax: +98 21 88991117; E-mail: m asadi@razi.tums.ac.ir

hepatitis in endemic and industrialized countries<sup>11</sup>.

There are a lot of techniques used for studying of these viruses, herein we focused on phage display technique used in hepatitis researches though.

### Literature search

Articles were searched from Google Scholar and Pubmed with key words of phage display and hepatitis viruses, Phage display and BAV, HBV, HCV. The valued information was subjected for review.

### Phage display technique

Phage display, created by G. Smith in 1985, describes an in vitro selection method in which a peptide or protein is expressed as a fusion with a coat protein of a bacteriophage, resulting in display of the fused protein on the surface of the phage particle, while the DNA encoding the fusion resides within the phage virion. Phage display has been used to generate a physical linkage between vast libraries of random peptide sequences to the DNA encoding each sequence, allowing rapid detection of peptide ligands for a variety of target molecules (antibodies, enzymes, cell-surface receptors, etc.) by an *in vitro* selection process called biopanning<sup>12-13</sup>. In its simplest form, biopanning is carried out by incubating a library of phage-displayed peptides with a plate coated with the target, washing away the unbound phage, and eluting the specifically-bound phage. The eluted phages are then amplified and taken through additional cycles of biopanning and amplification to successively enrich the pool of phage in favor of the tightest binding sequences. After 3-4 rounds, individual clones are characterized by DNA sequencing and ELISA.

Random peptide libraries displayed on phage have been used in a number of applications<sup>13</sup>, including epitope mapping<sup>14-16</sup>, mapping protein-protein contacts<sup>17</sup>, and identification of peptide mimics of non-peptide ligands<sup>18-22</sup>. Bioactive peptides have been identified either by panning against immobilized purifed receptors<sup>23</sup> or against intact cells<sup>24-26</sup>. The most common bacteriophages used in phage display are E.coli filamentous bacteriophages (f1, fd, M13)<sup>27-28</sup>,thoughT4, T7, and » phage have also been used<sup>29-31</sup>.

## Phage display technique applications

Applications of phage display

technology include determination of interaction partners of a protein (which would be used as the immobilized phage "bait" with a DNA library consisting of all coding sequence of a cell, tissue or organism) so that the function or the mechanism of the function of that protein may be determined<sup>32</sup>. Phage display is also a widely used method for in vitro protein evolution, also called protein engineering. As such, phage display is a useful tool in drug discovery. It is used for finding new ligands (enzyme inhibitors, receptor agonists and antagonists) to target proteins<sup>33-35</sup>. The technique is also used to determine tumor antigens (for use in diagnosis and therapeutic targeting)<sup>36</sup> and in searching for protein-DNA interactions using specially-constructed DNA libraries with randomized segments<sup>37</sup>. Beside these, phage display technique has been used in hepatitis studies too. In this review we focused on some applications of phage display in hepatitis researches.

### Phage display in developing a TaqMan real-time immuno-PCR method for detection of hepatitis viruses

PD-IPCR has been proven to be a highly sensitive assay for the detection of Hantaan virus nucleocapsid protein, prion protein<sup>38</sup> and the IgG in multiple sclerosis<sup>39</sup>.

A TaqMan real-time detection assay based on the concept of phage display mediated immuno-PCR (PD-IPCR) for the detection of HBcAg has been developed by Monjezi et. al (2012). PD-IPCR combines the advantages of immuno-PCR (IPCR) and phage display technology.

Previously, a phage bearing a constrained peptide (C-WSFFSNI-C) which interacts tightly with HBcAg was isolated<sup>40</sup> and specificity study showed that the phage only reacted with HBcAg but did not react with HBsAg and HbeAg<sup>41</sup>. In this strategy this phage was used to establish a PD-IPCR as an alternative choice for diagnosis of HBcAg.

Detection of HBcAg in serum by this method sonsist of 5 steps: a) HBcAg particle separated from the virion and dissociation to HBcAg dimers according to Kimura protocol<sup>42</sup>. b) The obtained dimmers coated on a microtiter plate well. c) M13 phages displaying the sequence C-WSFFSNI-C which interacts tightly with HBcAg<sup>40</sup> were added and allowed to interact with HBcAg. d) In order to lyse the bound phages and to release their genome, the plates well were heated at 95° C.
e) the released genome of phage used as a template in the TaqMan real-time PCR<sup>43</sup>.

The established TaqMan based real-time PD-IPCR can detect 10 ng of HBcAg by using 108 pfu/ml of the recombinant phage so it's about 10,000-fold more sensitive than the phage-ELISA. Therefore, the suggested PD-IPCR method may be an alternative option for the detection of HBcAg in serum samples. Collectively, these results indicate that this novel test could be a sensitive and highly reproducible detection method in hepatitis studies.

# Phage display and finding antibodies against the hepatitis viruses

The significance of phage display in antibody production is increasing. Phage display of antibody libraries has provided a powerful tool for the isolation of human MAbs to important viral pathogens<sup>44-49</sup>. Various formats of antibodies can be displayed on the surface of filamentous phage particles (e.g., M13), and antibodies with desired specificity can be isolated by panning on the antigens of interest<sup>50-51</sup>.

To date, a lot of antibodies have been developed in hepatitis researches. Among the first obtained antibodies against the hepatitis viruses was the successful molecular cloning of the antibody repertoire from an HCV-positive patient and the subsequent isolation of genes coding for anti-HCV human antibody fragments in Plaisant study. Availability this combinatorial library was panned against HCV and sixteen human antibody Fab fragments able to bind to HCV-specific antigens were generated which majority of them appeared to have specificity for the HCV c33 peptide<sup>52</sup>.

Beside Fab fragment libraries, human single chain Fv antibody (scFv) phage display library against hepatitis C virus has been screened. Yan et. al panned a human single chain Fv (scFv) phage antibody library against hepatitis C virus E2 antigen. The identified antibody was successfully applied in immunohistochemistry staining<sup>53</sup>.

In two similar studies, an antibody antigen-binding fragment (Fab) phage display library generated from a donor chronically infected with HCV screened against HCV E2 glycoprotein and finally within a total of more than 120 clones, ten Fabs from different heavy-chain groups recognizing the five different antigenic regions were converted into full-length IgG1s. All recombinant mAbs bound the genotype 1a HCV E1-E2 complex with approximately similar apparent affinities, but only one of these mAbs reacted with genotype 2a HCV. This finding suggesting that the epitopes related to that antigenic part are highly conserved. So using phage display library it could be possible to find neutralizing antibodies protect against hepatitis viruses. Such results provide evidence that protection against heterologous viral infection is possible, suggesting prophylactic vaccine against hepatitis viruses may be achievable. The mAb panel may also be useful for probing the antigenicity of E1-E2-based HCV vaccine candidates and guide the design of immunogens to elicit cross-Nabs (neutralizing antibodies) to HCV<sup>54-55</sup>.

Such studies have been done for HBV and HEV too. In Gwang study, for in vitro selection of antibodies that neutralize HBV, a large nonimmunized human phage antibody library in scFv format panned and two anti-pre-S1 antibodies obtained. These antibodies may be a good candidate for immunoprophylaxis against HBV infection<sup>56</sup>.

Infected chimpanzee, the primate most closely related to humans, derived library has been used for antibody production too. In Schofield study 2 mAbs were obtained from a cDNA phage display library of chimpanzee against ORF2 protein of HEV. These antibodies could be used in western blot and ELISA methods, both of them neutralized HEV and injection of virus-antibody mixtures to rhesus monkeys prevented infection<sup>57</sup>.

These researches described in the above showed that it is possible to state that the development of combinatorial antibody libraries displayed on the surface of phage offers the possibility of accessing monoclonal antibodies specificities against hepatitis viruses. Moreover detailed information about different epitopes that appear to be protective and definition of conserved elements in the viral envelope can be of capital importance for rational vaccine and drug design<sup>58</sup>. **Ligand identification against hepatitis viruses by phage display method** 

Identification of ligand binding is very

usual in phage display studies<sup>59-62</sup>. Some researchers have been done for developing of ligand binding peptide against the hepatitis viruses. The first ligand isolation against core antigen of hepatitis B virus (HBcAg) was done by Murray e. al. They screened a random hexapeptide library displayed on filamentous phage and introduced the "ALLGRMK" sequence as a binding ligand against truncated HBcAg. This peptide may represent a lead antiviral agent in chronic infections<sup>63</sup>.

Ho et. al isolated 2 other peptides (WSFFSNI and WPFWGPW) form a cyclic (disulfide constrained) phage peptide library reacted with full length HBc Ag<sup>40</sup>.

In an existing study, a peptide ligand (LPVRPWT or CD1) against  $\beta$ -cyclodextrin ( $\beta$ -CD) beads was found. A fusion peptide composed of CD1 peptide and residues 9–21 of the HCV core protein C1 (CD1HCV peptide) was designed. This fusion peptide was immobilized on  $\beta$ -CD beads by CD1 peptide. Using enzyme immunoassay, detection of anti-HCV Ab performed. Their findings showed anti-HCV Ab could react strongly, with a detection limit of 1 ng, to HCV core protein C1 conjugated to  $\beta$ -CD via LPVRPWT ligand peptide<sup>64</sup>.

These summarized results shows that high affinity ligands related to hepatitis viruses could be obtained from phage display libraries. Such ligands may provide useful information for detection and designing smaller and more potent peptides or small molecules that may be used in hepatitis researches.

# Phage display function in finding mimotopes against hepatitis viruses

Phage-displayed peptide approach can be used to identify mimotope of hepatitis viruses. Mimotopes are macromolecules which mimic the structure of an epitope and are frequently obtained from phage display random peptide libraries through screening<sup>65-66</sup>.

Puntoriero et.al, in order to identify synthetic surrogates of the HVR1 able to induce antibodies that reacted with virtually all HCV HVR1 variants, derived a consensus profile from more than 200 hyper variable region 1 (HVR1) sequences of different viral isolates and constructed a vast repertoire of synthetic HVR1 surrogates displayed on M13 bacteriophage as fusion to the major coat protein (pVIII). This library was panned against sera from clinically characterized HCV-infected individuals and some effective antigenic and immunogenic mimotopes of a large number of naturally occurring HCV variants isolated, most of which reacted with antibodies present in the majority of the sera from HCV-infected viremic patients (up to 80% of the tested samples). These findings are in good agreement with the hypothesis that HVR1 mimotopes have a higher capability of interacting with different anti-HVR1 antibodies than natural HVR1 variants<sup>67</sup>.

In another study a phage display library panned against anti human CD81 (hCD81) molecule (a putative receptor for HCV) and the peptide sequence ATWVCGPCT introduced as hCD81-like small peptide, which can block the binding site of HCV E2 for hCD81<sup>68</sup>.

Using the same procedure some mimtopes was isolated against HAV. These mimotopes mimics a structure of VP1 and VP3 capsid proteins. These peptides could bind specifically to serum antibodies from convalescent patients and immunization of mice by them resulted in neutralizing antibodies against HAV<sup>69</sup>.

Such finding suggest that such mimotpes could be have great potential for the development of synthetic peptide vaccines, DNA vaccines or could be used to develop a diagnostic assay for hepatitis viruses.

#### Phage as vaccine against hepatitis viruses

Potential application of phages as a modern platform for vaccines is another advantages of phages. It has been shown that phage could be use as antigen carriers in vaccine design strategies<sup>70-73</sup>.

Wan et. al constructed a filamentous phage particles that displayed the Hepatitis B virus epitope  $S_{28-39}$ . Injection of such phages to mice resulted in an MHC class I restricted HBs specific CTL response. These results suggest that the phage displaying desired peptide could be potentially suitable for developing the antihepatitis viruses vaccine studies<sup>74</sup>.

### CONCLUSION

The evaluated studies showed that phage display technique could play several roles in hepatitis studies. The function of this octopus like method, which its arms penetrated into different parts of biology, in hepatitis studies could be very variable. However, it looks that antibody and ligand studies share more concerns in this field of research.

#### REFERENCES

- Bernal, W. and J. Wendon, Acute liver failure. New England Journal of Medicine, 2013; 369(26): p. 2525-2534.
- Hens, N., et al., Model based estimates of longterm persistence of inactivated hepatitis A vaccine-induced antibodies in adults. Vaccine, 2014. **32**(13): p. 1507-1513.
- Hruska, J.F. and W.S. Robinson, The proteins of hepatitis B Dane particle cores. *Journal of medical virology*, 1977. 1(2): p. 119-131.
- World Health Organization. Hepatitis B. 2008. . [Online] Available from: http://www.who.int/ mediacentre/factsheets/ fs204/en/.
- 5. Everson, G.T., Living with hepatitis C: A survivor's guide. 2009: Hatherleigh Press.
- Choo, Q.-L., et al., Isolation of a cDNA clone derived from a blood-borne non-A, non-B viral hepatitis genome. Science, 1989. 244(4902): p. 359-362.
- Alter, H.J., Chronic consequences of non-A, non-B hepatitis, in Current perspectives in hepatology. 1989, *Springer*. p. 83-97.
- Rizzetto, M., The delta agent. Hepatology, 1983. 3(5): p. 729-737.
- 9. Farci, P., Delta hepatitis: an update. Journal of hepatology, 2003. 39: p. 212-219.
- HADZIYANNIS, S.J., Review: hepatitis delta. Journal of gastroenterology and hepatology, 1997. 12(4): p. 289-298.
- 11. Mushahwar, I.K., Hepatitis E virus: molecular virology, clinical features, diagnosis, transmission, epidemiology, and prevention. Journal of medical virology, 2008. 80(4): p. 646-658.
- Parmley, S.F. and G.P. Smith, Antibodyselectable filamentous fd phage vectors: affinity purification of target genes. Gene, 1988. 73(2): p. 305-318.
- Cortese, R., et al., Identification of biologically active peptides using random libraries displayed on phage. Current opinion in biotechnology, 1995. 6(1): p. 73-80.
- Scott, J.K. and G.P. Smith, Searching for peptide ligands with an epitope library. Science, 1990. 249(4967): p. 386-390.
- Cwirla, S.E., et al., Peptides on phage: a vast library of peptides for identifying ligands. Proceedings of the National Academy of

Sciences, 1990. 87(16): p. 6378-6382.

- Felici, F., et al., Selection of antibody ligands from a large library of oligopeptides expressed on a multivalent exposition vector. Journal of molecular biology, 1991. 222(2): p. 301-310.
- 17. Hong, S.S. and P. Boulanger, Protein ligands of the human adenovirus type 2 outer capsid identified by biopanning of a phage-displayed peptide library on separate domains of wildtype and mutant penton capsomers. The EMBO journal, 1995. 14(19): p. 4714.
- Devlin, J.J., L.C. Panganiban, and P.E. Devlin, Random peptide libraries: a source of specific protein binding molecules. Science, 1990. 249(4967): p. 404-406.
- Oldenburg, K.R., et al., Peptide ligands for a sugar-binding protein isolated from a random peptide library. Proceedings of the National Academy of Sciences, 1992. 89(12): p. 5393-5397.
- Scott, J.K., et al., A family of concanavalin Abinding peptides from a hexapeptide epitope library. Proceedings of the National Academy of Sciences, 1992. 89(12): p. 5398-5402.
- Hoess, R., et al., Identification of a peptide which binds to the carbohydrate-specific monoclonal antibody B3. Gene, 1993. 128(1): p. 43-49.
- O'Neil, K.T., et al., Identification of novel peptide antagonists for GPIIb/IIIa from a conformationally constrained phage peptide library. Proteins: Structure, Function, and Bioinformatics, 1992. 14(4): p. 509-515.
- Doorbar, J. and G. Winter, Isolation of a peptide antagonist to the thrombin receptor using phage display. Journal of molecular biology, 1994. 244(4): p. 361-369.
- Goodson, R.J., et al., High-affinity urokinase receptor antagonists identified with bacteriophage peptide display. Proceedings of the National Academy of Sciences, 1994. 91(15): p. 7129-7133.
- Barry, M.A., W.J. Dower, and S.A. Johnston, Toward cell-targeting gene therapy vectors: Selection of cell-binding peptides from random peptide-presenting phage libraries. Nature medicine, 1996. 2(3): p. 299-305.
- McLafferty, M., et al., M13 bacteriophage displaying disulfide-constrained microproteins. Gene, 1993. 128(1): p. 29-36.
- Kehoe, J.W. and B.K. Kay, Filamentous phage display in the new millennium. Chemical reviews, 2005. 105(11): p. 4056-4072.
- 28. Smith, G.P. and V.A. Petrenko, Phage display. Chemical reviews, 1997. 97(2): p. 391-410.
- 29. Malys, N., et al., A bipartite bacteriophage T4 SOC and HOC randomized peptide display

library: detection and analysis of phage T4 terminase (gp17) and late ó factor (gp55) interaction. Journal of molecular biology, 2002. 319(2): p. 289-304.

- Danner, S. and J.G. Belasco, T7 phage display: a novel genetic selection system for cloning RNA-binding proteins from cDNA libraries. Proceedings of the National Academy of Sciences, 2001. 98(23): p. 12954-12959.
- 31. Kalniòa, Z., et al., Evaluation of T7 and lambda phage display systems for survey of autoantibody profiles in cancer patients. Journal of immunological methods, 2008. 334(1): p. 37-50.
- 32. Arap, W., et al., Steps toward mapping the human vasculature by phage display. Nature medicine, 2002. 8(2): p. 121-127.
- Lunder, M., et al., Comparison of bacterial and phage display peptide libraries in search of target-binding motif. Applied biochemistry and biotechnology, 2005. 127(2): p. 125-131.
- Bratkoviè, T., et al., Affinity selection to papain yields potent peptide inhibitors of cathepsins L, B, H, and K. Biochemical and biophysical research communications, 2005. 332(3): p. 897-903.
- Lunder, M., et al., Peptide inhibitor of pancreatic lipase selected by phage display using different elution strategies. Journal of lipid research, 2005. 46(7): p. 1512-1516.
- 36. Hufton, S.E., et al., Phage display of cDNA repertoires: the pVI display system and its applications for the selection of immunogenic ligands. Journal of immunological methods, 1999. 231(1): p. 39-51.
- Gommans, W.M., H.J. Haisma, and M.G. Rots, Engineering zinc finger protein transcription factors: the therapeutic relevance of switching endogenous gene expression on or off at command. Journal of molecular biology, 2005. 354(3): p. 507-519.
- Guo, Y.-C., et al., Phage display mediated immuno-PCR. Nucleic acids research, 2006. 34(8): p. e62-e62.
- Yu, X., et al., Characterization of phage peptide interaction with antibody using phage mediated immuno-PCR. Journal of immunological methods, 2007. 326(1): p. 33-40.
- Ho, K.L., et al., Selection of high affinity ligands to hepatitis B core antigen from a phage displayed cyclic peptide library. Journal of medical virology, 2003. 69(1): p. 27-32.
- 41. Hasmoni, S.S., Detection of hepatitis B core antigen using a fusion bacteriophage. 2005, Universiti Putra Malaysia.
- 42. Kimura, T., et al., New enzyme immunoassay

for detection of hepatitis B virus core antigen (HBcAg) and relation between levels of HBcAg and HBV DNA. Journal of clinical microbiology, 2003. 41(5): p. 1901-1906.

- 43. Monjezi, R., et al., Detection of hepatitis B virus core antigen by phage display mediated TaqMan real-time immuno-PCR. Journal of virological methods, 2013. 187(1): p. 121-126.
- 44. Bender, E., et al., Recombinant human antibodies: linkage of an Fab fragment from a combinatorial library to an Fc fragment for expression in mammalian cell culture. Human Antibodies, 1993. 4(2): p. 74-79.
- 45. Burton, D.R., et al., A large array of human monoclonal antibodies to type 1 human immunodeficiency virus from combinatorial libraries of asymptomatic seropositive individuals. Proceedings of the National Academy of Sciences, 1991. 88(22): p. 10134-10137.
- 46. Crowe, J.E., et al., Recombinant human respiratory syncytial virus (RSV) monoclonal antibody Fab is effective therapeutically when introduced directly into the lungs of RSV-infected mice. Proceedings of the National Academy of Sciences, 1994. 91(4): p. 1386-1390.
- Ditzel, H.J., et al., Mapping the protein surface of human immunodeficiency virus type 1 gp120 using human monoclonal antibodies from phage display libraries. Journal of molecular biology, 1997. 267(3): p. 684-695.
- 48. Geoffroy, F., R. Sodoyer, and L. Aujame, A new phage display system to construct multicombinatorial libraries of very large antibody repertoires. Gene, 1994. 151(1): p. 109-113.
- 49. Thompson, J., et al., Affinity maturation of a high-affinity human monoclonal antibody against the third hypervariable loop of human immunodeficiency virus: use of phage display to improve affinity and broaden strain reactivity. Journal of molecular biology, 1996. 256(1): p. 77-88.
- Fernández, L.A., Prokaryotic expression of antibodies and affibodies. Current opinion in biotechnology, 2004. 15(4): p. 364-373.
- 51. Mahaffey, K.W., et al., Effect of Pexelizumab, an Anti-C5 Complement Antibody, as Adjunctive Therapy to Fibrinolysis in Acute Myocardial Infarction The COMPlement inhibition in myocardial infarction treated with thromboLYtics (COMPLY) Trial. Circulation, 2003. 108(10): p. 1176-1183.
- 52. Plaisant, P., et al., Human monoclonal recombinant Fabs specific for HCV antigens obtained by repertoire cloning in phage display

combinatorial vectors. Research in virology, 1997. 148(2): p. 165-169.

- 53. Zhong, Y.-W., et al., Preparation of human single chain Fv antibody against hepatitis C virus E2 protein and its identification in immunohistochemistry. World journal of gastroenterology, 2002. 8(5): p. 863.
- 54. Law, M., et al., Broadly neutralizing antibodies protect against hepatitis C virus quasispecies challenge. Nature medicine, 2008. 14(1): p. 25-27.
- 55. Giang, E., et al., Human broadly neutralizing antibodies to the envelope glycoprotein complex of hepatitis C virus. Proceedings of the National Academy of Sciences, 2012. 109(16): p. 6205-6210.
- Park, S.-G., et al., Hepatitis B virus-neutralizing anti-pre-S1 human antibody fragments from large naïve antibody phage library. Antiviral research, 2005. 68(3): p. 109-115.
- 57. Schofield, D., et al., Identification by phage display and characterization of two neutralizing chimpanzee monoclonal antibodies to the hepatitis E virus capsid protein. Journal of virology, 2000. 74(12): p. 5548-5555.
- Dormitzer, P.R., J.B. Ulmer, and R. Rappuoli, Structure-based antigen design: a strategy for next generation vaccines. Trends in biotechnology, 2008. 26(12): p. 659-667.
- 59. Kahle, J., et al., Epitope mapping via selection of anti-FVIII antibody-specific phage-presented peptide ligands that mimic the antibody binding sites. Thrombosis and haemostasis, 2015. 113(2): p. 396-405.
- Liu, G.W., et al., Efficient identification of murine M2 macrophage peptide targeting ligands by phage display and next-generation sequencing. Bioconjugate chemistry, 2015. 26(8): p. 1811-1817.
- Gunay, K.A. and H.-A. Klok, Identification of Soft Matter Binding Peptide Ligands Using Phage Display. Bioconjugate chemistry, 2015. 26(10): p. 2002-2015.
- 62. Zhang, D., et al., Screening and Identification of a Phage Display Derived Peptide That Specifically Binds to the CD44 Protein Region Encoded by Variable Exons. Journal of biomolecular screening, 2016. 21(1): p. 44-53.
- 63. Dyson, M.R. and K. Murray, Selection of peptide inhibitors of interactions involved in complex protein assemblies: association of the core and surface antigens of hepatitis B virus.

Proceedings of the National Academy of Sciences, 1995. 92(6): p. 2194-2198.

- 64. Kang, B. and S.-J. Choi, Identification of a polymeric â-cyclodextrin-binding peptide from a phage-displayed peptide library. Analytical biochemistry, 2011. 415(1): p. 46-51.
- Collins, J., Phage display, in Annual reports in combinatorial chemistry and molecular diversity. 1997, Springer. p. 210-262.
- 66. Asadi-Ghalehni, M., et al., in Silico and in Vitro Evaluation of A Recombinant Fusion Peptide as A Novel Candidate Vaccine for EGFR-Positive Tumors. Biosciences Biotechnology Research Asia, 2015. 12(3): p. 2405-2410.
- 67. Puntoriero, G., et al., Towards a solution for hepatitis C virus hypervariability: mimotopes of the hypervariable region 1 can induce antibodies cross reacting with a large number of viral variants. The EMBO journal, 1998. 17(13): p. 3521-3533.
- Cao, J., et al., Selection of a phage-displayed peptide recognized by monoclonal antibody directed blocking the site of hepatitis C virus E2 for human CD81. Journal of microbiological methods, 2007. 68(3): p. 601-604.
- Larralde, O.G., et al., Identification of hepatitis A virus mimotopes by phage display, antigenicity and immunogenicity. Journal of virological methods, 2007. 140(1): p. 49-58.
- Asadi-Ghalehni, M., et al., Cancer immunotherapy by a recombinant phage vaccine displaying EGFR mimotope: an in vivo study. Immunopharmacology and immunotoxicology, 2015. 37(3): p. 274-279.
- Javanmardi, M., et al., Triple tandem mimotope peptide of Epidermal Growth Factor Receptor displaying on the surface of M13 phage induces anti-tumor response in mice tumor model. Iranian Journal of Biotechnology, 2014. 12(3): p. 9-17.
- 72. Van Houten, N., et al., Filamentous phage as an immunogenic carrier to elicit focused antibody responses against a synthetic peptide. Vaccine, 2006. 24(19): p. 4188-4200.
- van Houten, N.E., et al., Engineering filamentous phage carriers to improve focusing of antibody responses against peptides. Vaccine, 2010. 28(10): p. 2174-2185.
- Wan, Y., et al., Induction of hepatitis B virusspecific cytotoxic T lymphocytes response in vivo by filamentous phage display vaccine. Vaccine, 2001. 19(20): p. 2918-2923.